## **Biospin Plasmid DNA Mini Extraction Kit**

#### TECHNICAL SUPPORT:

For technical support, please dial phone number : 0086-571-87774567-5287 or 5297, or fax to 0086-571-87774553

Email to reagent@bioer.com.cn.

Website: www.bioer.com.cn

#### **Kit Components**

Cat#	BSC01S1	BSC01M1	BSC01L1	
Component	50T	100T	250T	
Resuspension Buffer	12.5 mL	25 mL	62.5 mL	
Lysis Buffer	12.5 mL	25 mL	62.5 mL	
Neutralization Buffer	17.5 mL	35 mL	87.5 mL	
Wash Buffer	15 mL (add 60 mL	15 mL×2 (add 60 mL	37.5 mL×2 (add 150	
	ethanol before use)	ethanol before use)	mL ethanol before use)	
Elution Buffer	10 mL	20 mL	50 mL	
RNase A	50μL	100μL	250μL	
Spin Column	50 tubes	100 tubes 250 tubes		
Handbook	1сору	1copy	1copy	

## Storage and transportation

- ♦ The kit should be stored at room temperature(15~25°C), but the RNase solution should be stored at 2~8°C. The kit can be stored for up to 18 months by this method. After addition of RNase solution, Resuspension Buffer should be stored 2~8°C.
- ◆ The kit can be transported at room temperature.

#### Introduction

The kit provides a fast, simple, and cost-effective plasmid miniprep method for routine molecular biology laboratory applications. Plasmid DNA can be purified from 1-5mL of overnight cultures of E. coli. The DNA isolated by this kit is ready for downstream applications such as restriction enzyme digestion, sequencing, PCR/Real-time PCR and other downstream experiments.

#### **Technical Information**

Method	Work time	Column volume	Column yield	Elution recovery	Plasmid DNA length	Culture volume			
Spin column	25 min for 24 samples	750 µL	20 μg DNA	≥99%	≤10kB	1~5mL high-copy plasmid			

## Apparatus and materials to be prepared by the user

## **Important notes**

- 1. The RNase solution should be all added into the Resuspension Buffer before use, mix and store at 2-8°C.
- 2. Add 60mL absolute ethanol to Wash Buffer before use and mix well.
- 3. If the Lysis Buffer and Neutralization Buffer precipitated, it should be redissolve by warming to 37 °C. Please not vortex Lysis Buffer acutely.
- 4. Please close the lid immediately after using Lysis Buffer so as to avoid acidification.
- 5. The kit can extract high-quality plasmid DNA from 1-5mL E.coli overnight cultured.
- 6. The suitable volume is 50 µL for Elution Buffer; user can adjust its volume if necessary.

#### **Procedure**

- 1. Add 1-1.5mL cultured bacteria to 1.5mL micro centrifuge tube.
- 2. centrifuge at 10, 000rpm  $(8,000 \sim 10,000 \times g)$  for 30s, and discard the supernatant. Step 1th and 2th could be repeated for more than one time to collect enough cells.
- 3. Resuspend pelleted bacterial cells in 250 µL Resuspension Buffer and No cell clumps should be visible after resuspension of the pellets.
- 4. Add 250 µL Lysis Buffer and gently invert the tube 4-6 times to mix. Do not vortex, as this will result in shearing of genomic DNA. Do not allow this step for more than 5 minutes.
- 5. Add 350 µL Neutralization Buffer and gently invert the tube 4-6 times to mix. The solution should become cloudy and no local precipitate should be visible.
- 6. Centrifuge for 10minutes at 13,000rpm ( $>14,000 \times g$ ) until a compact white pellet form.
- 7. Apply the supernatant to the Spin column and centrifuge for 30-60 seconds at  $6,000 \times g$ . Discard the flow-through.
- 8. Add 650  $\mu$ L Wash Buffer to the Spin column and Centrifuge for 30-60 seconds at 12,000×g. Discard the flow-through.
- 9. Repeat step 8th once.
- 10. Centrifuge for an additional 1 minute at  $12,000 \times g$  and transfer the Spin column to a sterile 1.5mL micro centrifuge tube. Recommend to centrifuge according to this step; otherwise, there will be residual liquid in the column.
- 11. Add  $50\,\mu\text{L}$  Elution Buffer, ddH<sub>2</sub>O or TE Buffer to the Spin column and let it stand for 1 minute at room temperature. The volume of elution buffer could be adjusted according to needs.
- 12. Centrifuge for 1 minute at  $12,000 \times g$ . The buffer in the micro centrifuge tube contains the plasmid DNA.
- 13. The purified plasmid DNA can be used directly for kinds of downstream molecular biological experiments. Store at -20°C if not used immediately.

## **Troubleshooting**

## Get on plasmid DNA

If the plasmid DNA is not found in elution buffer, please check whether the ethanol had been added to Wash Buffer according to the volume be marked on bottle label.

## Low plasmid DNA yields

- 1) Please make sure that the bacteria was cultured in the right way.
- 2) The bacteria cells should be resuspented completely.
- 3) Incubate the Elution Buffer in  $30\sim60^{\circ}$ C, it will increase the yields.

## Absorbance problem

- 1) Absorbance is the difference from sample and criterion, please use the Elution Buffer to adjust to zero value and dilute the sample.
- 2) If the ratio of OD260/ OD230 is low, wash the spin column for one more time.
- 3) In the case of low ratio of OD260-320/ OD280-320, there is protein contamination, please add Neutralization Buffer, and then centrifuge buffer with sufficient rotating speed, thus to make precipitation compact; be careful to pipette supernatant so as to avoid pipette precipitation.
- 4) If the ratio of OD260-320/OD280-320 is high, add RNase A to Resuspension Buffer (100 $\mu$ g/mL).

## **Electrophoresis problem**

- 1) If there is genomic DNA in the result. Please invert the tube gently (step4 and 5).
- 2) If there is RNA in the result, add RNase A to Resuspension Buffer (100µg/mL).

## **DNA** Analysis

**Absorbance** analysis

# Get some plasmid DNA, diluted in a advisable factor with elution buffer. Survey the OD260, OD280 and OD320

Expressions: concentration ( $\mu$ g/mL) =50×OD260×dilution fact

Target: 2.0\(\geq\)OD260-320/ OD280-320\(\geq\)1.8

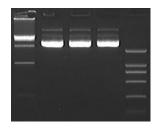
Notice: 1.0≥OD260≥0.1, the result of ratio is much reliable.

### **Agarose Gel Analysis**

## 0.8~1% Agarose gel

Example 1: Plasmid DNA electrophoresis

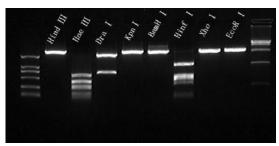
DL15, 000 DL2, 000



Example 2: Enzymatic reactions analysis

DL2,000

DL15,000



## **Company Information**

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